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# Free radical scavenging activity of methanolic leaf extract of *Pimenta dioica* on streptozotocin-induced diabetic rats

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#### Abstract

The present study was designed to investigate the free radical scavenging activity of *Pimenta diocia* on streptozotocin (STZ)-induced diabetic rats. The methanolic leaf extract of *Pimenta diocia* at the doses of 75 and 150 mg/ kg of body weight was administered orally once in a day to the diabetic induced group for 45 days. Glibenclamide (0.6 mg/kg of body weight) was used as reference drug. The antioxidant properties were assessed by estimating the liver and kidney catalase (CAT), thiobarbituric acid reactive substances (TBARS), superoxide dismutase (SOD), glutathione peroxidase (Gpx) and reduced glutathione (GSH). Antioxidant levels were significantly restored towards normal levels in *P.dioica* treated rats when compared with the STZ control. The results of the study indicate that the *Pimenta dioica* leaf methanolic extract exhibit promising antioxidant activity towards diabetic rats.

**Key words:** Free radical scavenging activity, *Pimenta dioica*, glibenclamide, catalase.

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#### 1. Introduction

Diabetes mellitus (DM) is a chronic disease caused by inherited or acquired deficiency in the production of insulin by the pancreas or by the ineffectiveness of the produced insulin. Such a deficiency results in increased concentrations of glucose in the blood, which in turn damage many of the body systems and in particular the blood and nervous systems. It is one of the alarming worldwide health present problems at leading to microvascular (retinopathy, neuropathy and nephropathy) and macrovascular

(heart attack, stroke and peripheral vascular disease) complications [1]. It is expected that about 366 million people are likely to be diabetic by the year 2030 [2]. Hyperglycemia is known to produce reactive oxygen species (ROS) which plays a central role in the complications of diabetes [3]. Diabetes is associated with oxidative stress, leading to an increased production of reactive oxygen species (ROS), including superoxide radical, hydrogen peroxide and hydroxyl radicals or reduction of antioxidant defense system. Implications of oxidative stress in the pathogenesis of diabetes is suggested not only by oxygen free radical generation but also due to non enzymatic protein glycosylation, auto oxidation of glucose, impaired antioxidant enzymes and formation of peroxides. Lipid peroxidation is a key marker of oxidative stress that results in extensive membrane damage and dysfunction [4].

Treatment of diabetes with sulphonylureas biguanides and is associated with adverse side effects. However, complementary medicine has grown in popularity in recent years owing to its minimal side effects and appropriate action. Dietary measures and traditional therapies plant as prescribed bv Ayurvedic and other indigenous systems of medicine are used commonly in India. Many indigenous Indian medicinal plants have been found to be useful in the successful management of diabetes and some of them have been tested for their active ingredients. The World Health Organization (WHO) has also recommended the evaluation of the plant's effectiveness and conditions against chronic ailment in place of chemically synthesized drugs. Despite the development of new drugs and their validation by scientific criteria, research still continues in scientific community around the world to evaluate antidiabetic activities of raw plant materials or isolated natural products without adverse effects.

*Pimenta dioica* (L.) Merril (Family: Myrtaceae) is commonly known as Allspice in culinary. It takes its name from the aroma of dried berries, which smells like the combination of spices, especially cinnamon, cloves, ginger and nutmeg. Allspice owes its characteristic odour due to the presence of essential oil in the pericarp of the seeds. The plant Allspice is mentioned in the Wealth of India [5]. The natives of Kerala and Mangalore use Allspice leaves as medicine for pain, arthritis, fever and stress. The drug has derived the name "Allspice" since its aroma resembles the aroma of spices such as clove, nutmeg and cinnamon [6]. In India, the leaves of *Pimenta* are used to flavor rice which gives it a typical aroma. Allspice is considered as a very important spice in the meat industry which utilizes the powder of the berries for the tenderizing of meat [7, 8].

previously In all the mentioned pathological conditions, oxidative stress is one of the causes, which trigged the momentum to explore the P. dioica leaf extract for in vitro antioxidant activity. Antioxidant and hepatoprotective activity in CCl<sub>4</sub> (Carbon Tetra Chloride) induced liver toxicity of all spice leaves had been reported earlier [9]. In the present study Pimenta dioica leaves were subjected to evaluate the antioxidant activities against diabetes.

## 2. Materials and Methods Chemicals

Streptozotocin (STZ) was purchased from Sigma–Chemical Co. Bangalore. All other chemicals and reagents used for this study were of analytical grade.

# Plant material

*Pimenta diocia* was collected from Kumuli, Kerala State, India.

# Preparation of extract

The *Pimenta dioca* leaves were dried at room temperature and then were powdered using dry grinder and passed through sieve. Hundred grams of *Pimenta dioica* were packed in a soxhlet apparatus and extracted with methanol. The methanolic extracts were concentrated on a rotary evaporator.

## Experimental animals

Male Wistar albino rats (150-200 g) were procured from Rajah Muthiah Medical College and Hospital, Annamalai University, Chidambaram, Tamilnadu. India, and were housed in polycarbonate cages in an animal room with 12 hours day - night cycle. The animals were allowed free access to tap water and standard laboratory rat food. The animal treatment and protocol employed were approved by the TAEC, Annamalai University (Registration Number -1084 /2014/CPCSEA)

## Induction of experimental diabetes

Diabetes was induced in the rats by intraperitoneal (I.P.) iniection of streptozotocin (STZ) at a dose of 55 mg/kg b.w dissolved in 0.1 M cold citrate buffer (pH = 4.5) [10]. The rats were allowed to drink 5% glucose solution overnight to overcome the drug- induced hypoglycemia. The blood glucose values above 250 mg/dl on the third day after streptozotocin injection were considered as diabetic rats. Then the treatment was started the fifth dav on after streptozotocin injection and it was considered as the first day of treatment.

# **Experimental design**

All animals were randomly divided into five groups with six animals in each group

- 1. Normal untreated rats
- 2. Diabetic control rats (STZ) (55 mg/kg bw).
- 3. Diabetic rats treated with methanolic extract of *Pimenta diocia* leaves (75 mg/kg of body weight)
- 4. Diabetic rats treated with methanolic extract of *Pimenta diocia* leaves (150 mg/kg of body weight)
- 5. Diabetic rats treated with standard drug, glibenclamide ( $0.6 \mu g/kg$  of body weight).

## Estimation of antioxidant parameters

Tissues (liver and kidney) were dissected out and washed immediately with ice cold saline to remove any blood. The antioxidant enzymes such as superoxide dismutase (SOD) [11], reduced glutathione (GSH) [12] catalase (CAT) [13]. thiobarbituric acid reactive substances (TBARS) [14] and glutathione peroxidase (GPx) [15] activity were estimated in the liver and kidney.

# Statistical analysis

All antioxidant data are expressed as mean  $\pm$  S.E. Statistical analysis was performed using one-way ANOVA followed by Tukey's multiple tests using SPSS (version 18) of computer software. In all cases, P-value of less than 0.05 was considered to be significant.

## 3. Results

The antioxidant enzymes such as TBARS, SOD, CAT, GSH, and GPx were analyzed in the liver and kidney of normal and STZ induced diabetic rats and treated with the methanolic leaf extracts of Pimenta dioica and the standard drug glibenclamide (Table 1). An increased level of TBARS was observed and the levels of superoxide dismutase (SOD), catalase (CAT), glutathione peroxidase (GPx) and reduced glutathione (GSH) were significantly (p<0.05) reduced in STZ induced diabetic rats. These adverse changes were reversed to near normal values in the methanolic leaf extract of *P. dioica* treated rats on par with the results obtained during the administration of the standard drug, glibenclamide.

## 4. Discussion

Antioxidants are substances or nutrients which can prevent or slow down the oxidative damage to the body. When the body cells use oxygen, they naturally produce free radicals (by-products) which can cause damage [16].

Organs	Group	Treatment	TBARS	SOD	САТ	GSH	GPx
Liver	Ι	Normal control	3.90±0.10	112.90±2.28	32.55±0.90	241.28±3.40	34.50±0.81
	II	Diabetic control	8.41±0.16	81.01±1.48	21.60±0.31	220.35±1.32	22.10±0.39
	III	Diabetic + <i>P. dioica</i> extract (75 mg/kg b.w)	4.60±0.11	96.39±2.19	26.20±0.51	235.89±3.10	29.45±0.89
	IV	Diabetic + <i>P. dioica</i> extract (150 mg/kg b.w)	4.54±0.15	102.48±3.12	28.21±0.51	237.41±0.51	32.04±0.44
	V	Diabetic + glibenclamide (0.6 mg/kg b.w)	4.15±0.09	109.69±3.64	31.47±0.51	240.33±2.31	33.01±0.55
Kidney	Ι	Normal control	5.13±0.10	146.69±2.01	28.81±0.55	334.98±5.71	32.60±0.89
	II	Diabetic control	7.35±0.15	115.59±2.05	20.01±0.84	302.18±3.13	23.80±0.68
	III	Diabetic + <i>P. dioica</i> extract (75 mg/kg b.w)	5.93±0.10	131.12±3.41	27.60±0.50	326.80±4.98	27.02±0.59
	IV	Diabetic + <i>P. dioica</i> extract (150 mg/kg b.w)	5.39±0.20	133.10±3.65	28.16±0.55	329.41±6.41	30.12±1.02
	V	Diabetic + glibenclamide (0.6 mg/kg b.w)	5.27±0.15	144.81±2.90	28.61±0.51	331.65±3.41	31.42±1.15

 Table 1. Effect of *P. dioica* on the liver and kidney antioxidant enzymes in STZ – induced diabetic rats

Values are expressed as mean $\pm$ S.E (n=6) and are significantly different at p<0.005 when compared with control groups.

Chronic hyperglycemia in diabetes leads to auto-oxidation of glucose, non-

enzymatic protein glycosylation, impaired glutathione metabolism, alteration in

antioxidant enzymes and formation of lipid peroxides. The above events accelerate the production of free radicals and weaken the antioxidant defense system. Hence, attention has been given to naturally occurring antioxidants that counteract the deleterious effects of reactive antioxidants. The increase in oxygen free radicals in diabetes could be primarily due to an increase in the blood glucose levels, which upon auto-oxidation generate free radicals. The increased susceptibility of the tissues of the diabetic animals may be due to the activation of the lipid peroxidation system. The possible source of oxidative stress in diabetes includes shifts in redox balance resulting from altered carbohydrate and metabolism lipid and increased generation of reactive oxygen species [17]. The antioxidant activity of *P.dioica* in liver and kidney was studied in diabetic rats. After the induction of diabetes by STZ, significantly (P<0.005) decreased levels of SOD, CAT, GPx, reduced GSH and increased level of TBARS in liver and kidney were observed compared to normal control rats. These altered above antioxidant levels were reversed significantly (P<0.005) to near normal levels after the administration of P. dioica 75 and 150 mg/kg dose and glibenclamide 0.6µg/kg dose compared to diabetic control rats. It is well known that CAT, SOD and GPx play an important role as protective enzymes against free radical formation in the tissues [18]. These adverse change were reversed to near normal values in the methanolic extract of P. dioica leaf treated rats. Recent studies have clearly demonstrated the importance of medicinal plants in the treatment of experimental diabetes, where oxidative stress induced apoptosis or  $\beta$  -cell death occur [19, 20]. Oral administration of Asparagus racemosus (EEAR) showed significant hypoglycemic effects against

STZ-induced diabetes in rats. The extract significantly lowered the levels of blood glucose and TBARS and significantly increased the levels of GSH, SOD and CAT [21, 22]. From the present study it could be concluded that the methanolic leaf extract of *P. dioica* possess potent antioxidant properties in STZ induced diabetic rats.

#### References

- Umar A, Ahmed QU, Muhammad BY, Dogarai BB and Soad SZ: Antihyperglycemic activity of the leaves of Tetracera scandens Linn. Merr. (Dilleniaceae) in alloxan induced diabetic rats. Journal of Ethnopharmacology 2010; 1: 140–145.
- 2. Wild SG, Roglic A, Green R, Sicree R and King H: Global prevalence of diabetes: estimated for the year 2000 and projection for 2030. Diabetes Care 2004; 5: 1047–1053.
- Dewanjee S, Das AK, Sahu R and Gangopadhyay M: Antidiabetic activity of Diospyros peregrine fruit: effect on hyperglycemia, hyperlipidemia and augmented oxidative stress in experimental type 2 diabetes. Food and Chemical Toxicology 2009; 47: 2679– 2685.
- 4. Parisa S: Pharmacol. Res.2007: 56: 261-266
- 5. The Wealth of India. A dictionary of Indian raw materials. Publications and information directorate, CSIR, New Delhi 1969: Vol-8: pp 58-9.
- 6. Neal MC: In Gardens of Hawai'i. Bernice P. Bishop museum special publication 40, Bishop Museum Press, Honolulu, HI 1969.
- 7. Seidemann J: *Pimenta* Lindl.-Allspice-Myrtaceae. World Spice Plants, Springer-Verlag, Heidelberg. 2005: 286-287.
- 8. Sharma R. Pimenta :Medicinal plants of India –An encyclopedia. Data Publishing House Delhi, New Delhi 2003.
- 9. Nayak Y, Abhilash D, Vijaynarayana K and Fernandes J: Antioxidant and hepatoprotective activity of *Pimenta*

*dioica* leaves extract. *J Cell* Tissue Res 2008: 8(3); 1571-1576.

- Bandaranayake WM: Bioactivities, bioactive compounds and chemical constituents of mangrove plants.Wetlands Ecology and Management 2002:10 ; pp. 421–452
- 11. Kakkar P, Das B, Viswanathan PN: A modified spectrophotometric assay of superoxide dismutase. Ind J Biochem Biophys 1984: 21;130-132
- 12. Ellman GC: Tissue sulflydryl groups. Arch Biochem Biophys 1959: 82; 70 – 77.
- 13. Sinha AK: Colorimetric assay of catalase. Anal Biochem. Jun 1972:47(2); 389-94.
- 14. Niehaus WG, Samuelson B: Formation of malondialdehyde from phospholipids arachidonate during microsomal lipid peroxidation. Eur J Biochem 1968: 6 ; 126-130
- 15. Rotruck JT, Pope AL, Ganther HE Swanson AB, Hafeman DG and Hoekstra WG: Selenium biological role as a component of glutathione peroxidase. Science 1973: 179; 588-590.

- 16. Khalil OA, Ramadan KS, Danial EN, Alnahdi HS and Ayaz NO: Antidiabetic activity of Rosmarinus officinalis and its relationship with the antioxidant property. Afr J Pharm Pharmacol 2012: 6; 1031e1036.
- 17. Laight DW, Carrier MJ, Anggard EE: Antioxidants, diabetes and endothelial dysfunction. Cardiovasc Res 2000: 47; 457e464.
- Oberly, WR, Buettner, RG: Role of superoxide dismutase in cancer. Cancer Research 1974: 35; 1141-1149.
- 19. Kinloch RA, Treherne JM, Furness LM and Hajimohamadreza I: Trends Pharmacol Sci 1999: 20; 35-42
- 20. Sandhya SL, Shewade Y and Bhonde R: J. Ethnopharmacol 2000:73; 71-79. [20]
- 21. Palanisamy A, Sorimuthu PS: Chemico-Biological Interactions. 2007: 165; 155-164.
- 22. Bavara JH, Narasimhacharya AVR: I; Fitoterapia 2008: 79; 328- 331.