

Research Article

In-Vitro Anti Breast Cancer Activity of *Syzygium Cumini* Against MCF-7 Cell Line

Gitanjali Tripathy^{*1}, Debasish Pradhan¹

¹University Department of Pharmaceutical Sciences, Utkal University, Vanivihar, Bhubaneswar-04

Abstract

In this study, different concentrations of the methanolic extract of fruit pulp of the plant *Syzygium cumini* was subjected to in-vitro cytotoxic activity study against MCF-7cells using the MTT assay. Percentage cell viability of cell lines were carried out by using Trypan blue dye exclusion technique MTT assay was used to evaluate the reduction of viability of cell cultures in the presence and absence of the extract. Cell viability was inhibited to different extents by different concentrations of the extract.

Key words: *Syzygium cumini*; cytotoxic activity; crude extracts; MTT.

***Corresponding Author: Gitanjali Tripathy,** University Department of Pharmaceutical Sciences, Utkal University, Vanivihar, Bhubaneswar-04.

1. Introduction

Natural products, including plants, animals and minerals have been the basis of treatment of human diseases [1, 2]. History of medicine dates back practically to the existence of human civilization. The current accepted modern medicine or allopathy has gradually developed over the years by scientific and observational efforts of scientists. However, the basis of its development remains rooted in traditional medicine and therapies. The history of medicine includes many ludicrous therapies. Nevertheless, ancient wisdom has been the basis of modern medicine and will remain as one important source of future medicine and therapeutics [3,4].

Several commonly used herbs have been identified by the National Cancer Institute possessing cancer-preventive as properties. Those include members of the Allium sp. [garlic, onions and chives]; members of the Labiatae family [basil, mints, oregano, rosemary, sage, and thyme]; members of the Zingiberaceae family [turmeric and ginger]; members of the Umbelliferae family (anise, caraway, celery, chervil, cilantro, coriander, cumin, dill, fennel, and parsley) [5]. In addition, many herbs contain a variety of phytosterols, flavonoids. triterpenes, saponins, and carotenoids, which have been shown from studies of legumes, fruit, be and vegetables to cancer chemoprotective [6].

Breast cancer is the most commonly occurring cancer in women, comprising almost one third of all malignancies in females. It is second only to lung cancer as a cause of cancer mortality and it is the leading cause of death for American women between the ages of 40 and 55 [7]. The lifetime risk of a woman developing invasive breast cancer is 12.6 % 2 one ut of 8 females in the United States will develop breast cancer at some point in her life [8]. Syzygium cumini (Family-Myrtaceae) is native to India and East Indies. It is commonly called as Jamu Koli in Odia; Black Plum, Java Plum in English and Jamun in Hindi. The plant possesses acetyl oleanolic acid, triterpenoids, ellagic acid, isoquercitin, quercetin, kaempferol and myricetin in different concentrations [9]. Most of these compounds have been reported to possess antioxidant and free

2. Materials and Methods

radical scavenging activities [10].

Collection of plant material and extraction

Ripe fruits of Syzygium cumini was collected from the forest part of Bhubaneswar hill area situated in the eastern part of India in the month of May and identified by Dr. S K Sahu, a taxonomist at Utkal University. VaniVihar, Odisha, and Dr. S P Panda, keeper. Regional Herbarium Plant Research Center, Bhubaneswar. Voucher were deposited specimens in the herbarium of the Department of Botany, Utkal University. The fruits were cut into small pieces and the pulp was separated from the seeds. The pulp was shade-dried, and milled. The coarsely powdered, shade dried fruit pulp of both the plants was first defatted with Petroleum ether using soxhlet apparatus. The extracts were concentrated using rotary evaporator to

get solid residue. The marc from the central compartment was removed, dried and extracted by exhaustive extraction with a series of solvents of increasing polarity with Soxhlet extractor was done [11]. The weight of the residue extracts recorded and percent vield were calculated. Solvents used with increasing polarity are petroleum ether, ethyl acetate, methanol and the scheme of extraction was depicted as a flow chart (Figure 1) below.

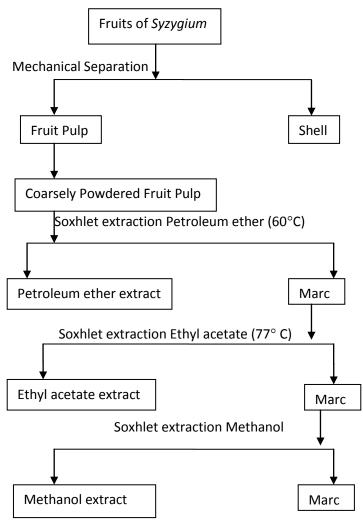


Figure 1: Scheme of extraction for the Fruit pulp of *Syzygium cumini*.

Preliminary Phytochemical Screening

The percentage yield of other extracts except methanolic extract was negligible. So, the methanolic extract was taken for further experimental work. The prepared methanolic extract was subjected to routine phytochemical analysis [12] to identify the presence of various phytochemicals such as carbohydrates, alkaloids, glycosides, saponins, flavonoids, tannins, sterols, phenols, etc.

Cell Culture

Human breast cancer MCF-7 cell line Sigma-Aldrich, was obtained from Bangalore. MCF-7 is a cell line used in many studies due to its characteristics and being easy to culture. The MCF-7 cell line is adherent and grows in clumps. The cells maintained were in **RPMI-1640** supplemented with 10% FBS, penicillin and streptomycin (100 U/ml), (100 μ g/ml) in a humidified atmosphere of 50 μ g/ml CO₂ at 37 °C.

In Vitro Anti-Breast Cancer Activity

Viability Staining by Trypan blue dye exclusion method Principle

Trypan Blue is a vital blue acid dye that has two azo chromophores group. The reactivity of Trypan blue was based on the fact that the chromophore was negatively charged and does not interact with the cell unless the membrane was damaged. Therefore, all the cells which exclude the dye are viable.

Trypan blue will not enter into the cell wall of plant cells grown in culture. It is used in estimating the number of viable cells present in a population.

Procedure

The Methanolic extract of *Syzygium cumini* (MESC 200 mg/kg) was studied for short term *in vitro* cytotoxicity using MCF-7cells.

From the above solutions 10mg of the extracts were taken in an Eppeandorf vial of capacity 1ml and diluted to six concentrations with different its duplicate and control (50%) using DMSO as a solvent and mixed with the help of a vortexing machine. The cell viability was checked by trypan blue dye (4%) [13,14]. The cell suspension $(2x10^6$ cells in 0.1ml) was added to tubes containing various concentrations of the test compounds and the standard drug Doxorubicin. The volume was made up to 1ml using Phosphate Buffered Saline (PBS).

Control tube contained only cell suspension. These assay mixtures were incubated for 3 hour at 37°C. After incubation 0.1 ml trypan blue was added and number of dead cells determined by using an Automated cell counter. The mixtures were examined for the viability of the cells (non - viable cells are stained and viable cells excluded the stain). Cells were counted by the following formulae:

Cell count = Number of cells ×Dilution (Area ×Thickness of fluid film)

Percentage of cell viability = (Live cell count/Total cell count) ×100

Micro culture tetrazolium (MTT) assay

Cell viability was assessed by MTT assay (Micro culture tetrazolium/ formazan assay) in the presence and absence of different concentrations of the plants extract. The cells were seeded in 96-well plates. Four wells for each concentration were seeded and triplicate plates were used the cell line. Then, the cells were incubated at 37° C. After 36 h of incubation, various concentrations of fruit extracts were added to the wells to obtain final concentrations of 62.2, 125, 250 and 1000 µg/ml, respectively. Control groups were mixed with DMSO to obtain a final

concentration of 1 %. Doxorubicin was used as positive control. The cells were incubated for an additional 48 h, 50 µl of [3-(4,5-dimethylthiazol-2-yl)-2,5-MTT diphenyl tetrazolium bromide] (1 mg/ml, Sigma) in phosphate buffered saline added to each well, and (PBS) was incubated for 4 h at 37 °C. The medium was removed and formazan was dissolved in DMSO and the optical density measured at 590 mm using a bioassay reader (Biorad, USA) [15].

 IC_{50} was defined as the concentration of the extract that produced a 50% decrease in cell viability relative to the negative control which was wells exposed to the solvent without any extract [16, 17].

Statistical Analysis

Data are reported as the mean \pm SD for at least three replicates. Statistical analysis was performed using the Student t-test, with significance level set at P < 0.05.

3. Results and Discussion

The various concentration of plant extract used were 62.5, 125, 250, 500 and 1000 μ g/ml, Doxorubicin and control (without extract). Decrease in cell count was

observed with increase in concentration of the extracts. There was a dose dependent increase in cytotoxic activity for all the concentrations tested. In vitro exposures of MCF-7 cells with various concentrations of Syzygium cumini extract (62.5, 125, 250, 500, 1000 µg/ml) significantly suppressed MCF-7 cancer cell growth dose-dependent in а manner (P<0.05). The maximum inhibition of MCF-7 cells due to exposure to MESC was found at 1000 μ g/ml of the extracts 70.12% inhibition. The was results showed dose dependent response against MCF-7 cell line. The cytotoxic activity may be due to the presence of secondary metabolites like flavonoids present in Svzvgium cumini.

The result of the present study reveals that, the *in vitro* anti breast cancer activity was observed in Trypan blue exclusion assay and MTT assay against MCF-7 cell line. Although the anti-breast cancer activity of Doxorubicin was more than that of the extracts, it may be noted that the drug is a single entity whereas the extract is a crude one containing numerous compounds.

Treatment and Conc. (mcg/ml)	Absorbance	% Inhibition	IC ₅₀	R ²
MESC (62.5)	0.427	14.25		
MESC (125)	0.380	23.69	266.8	0.9617
MESC (250)	0.268	46.05		
MESC (500)	0.165	66.73		
MESC (1000)	0.092	70.12		
Standard	0.00021	99.91		
Control	0.528			

Table 1: Determination of Cytotoxicity of MESC

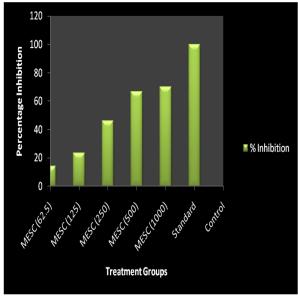


Figure 2 : Percentage growth inhibition of MESC, Control and Standard against MCF-7 cell line.

4. Conclusion

Studies have shown differential sensitivities to several natural compounds between tumor and normal cells in vitro or in vivo, and the results obtained from the present study show that the methanol extract from fruit pulp of Syzygium cumini had anti breast cancer activity against MCF-7 cell lines. Our phytochemical screening revealed the presence of flavonoids, alkaloids, steroids in the methanolic extract of Syzygium cumini, which could be responsible for this activity. Flavonoids have been found to possess antimutagenic and antimalignant effects [18, 19]. Moreover it has protective effect against breast cancer by their effect on signal transduction in cell proliferation and angiogenesis [20]. It also justifies the folklore medicinal uses and claims about the therapeutic values of this plant as curative agent against cancer and we therefore, suggest further. the purification and characterization of the phytochemicals along with investigations are needed to provide some additional

insight into the *in vivo* cytotoxic activity of the plants with a view to obtaining useful chemotherapeutic agent.

Acknowledgements

The authors would like to thank the Head of the Department, University Department of Pharmaceutical Sciences, Utkal University, Vani Vihar, Bhubaneswar, India for support in carrying out this work. The authors are thankful to INSPIRE-DST and UGC for providing financial support for the smooth conduct of research work.

References

- 1. Jacob, E. (2009). Natural Products-Based Drug Discovery: Some Bottlenecks and Considerations. *Current Science*, 96(6), pp.753-754.
- 2. Butler, M. (2004). The Role of Natural Product Chemistry in Drug Discovery. *J. Nat. Prod.*, 67(12), pp.2141-2153.
- 3. Dev, S. (1997). Ethnotherapeutic and modern drug development: The potential of Ayurveda.*Curr Sci.*, 73, pp.909-928.
- Newman, D., Cragg, G. and Snader, K. (2003). Natural Products as Sources of New Drugs over the Period 1981–2002. *J. Nat. Prod.*, 66(7), pp.1022-1037.
- Chan, T., Chan, J., Tomlinson, B. and Critchley, J. (1993). Chinese herbal medicines revisited: a Hong Kong perspective. *The Lancet*, 342(8886-8887), pp.1532-1534.
- 6. Nishimura, H., Ariga, T., Huang, M., Osawa, T., Ho, C. and Rosen, R. (1993). Food Phytochemicals for Cancer Prevention I. *ACS Symposium Series*, pp.128-143
- 7. Harris, J., Lippman, M., Veronesi, M. and Willet, W. (2015). Breast cancer. *N Engl J Med*, 327, pp.319-328.
- 8. Greenlee, R., Hill-Harmon, M., Murray, T. and Thun, M. (2001). Cancer statistics, 2001. *CA Cancer J Clin.*, 51(1), pp.15-36.
- 9. Rastogi, R. and Mehrotra, B. (1990). *Compendium of Indian Medicinal Plants*. Lucknow, India: Central Drug Research Institute, pp.388-389.

- 10. Tanaka, M., Chiu, W., Nagashima, Y. and Taguchi, T. (1988). Application of antioxidative Maillard reaction products from histidine and glucose to sardine products. *NIPPON SUISAN GAKKAISHI*, 54(8), pp.1409-1414.
- 11. Kokate, C. (1994). *Practical Pharmacognosy.* New Delhi: Vallabh Prakashan, p.135.
- 12. Harbone, J. (1998). *Phytochemical Methods, A guide to Modern Technique of plant analysis.,London, pp. 91.* London: Chapman and Hall Ltd., p.91.
- Dongre, S., Badami, S., Natesan, S. and Raghu, C. (2015). Antitumor Activity of the Methanol Extract of Hypericum hookerianum Stem Against Ehrlich Ascites Carcinoma in Swiss Albino Mice. J Pharmacol Sci, 103(4), pp.354-359.
- Moldeus, P., Hogberg, J., Orrhenius, S. and Parker, L. (1978). *Methods in enzymology*. 2nd ed. New York: Academic Press, pp.60-71.
- 15. Rubinstein, L., Shoemaker, R., Paull, K., Simon, R., Tosini, S., Skehan, P., Scudiero, D., Monks, A. and Boyd, M. (1990). Comparison of In Vitro Anticancer-Drug-Screening Data Generated With a Tetrazolium Assay Versus a Protein Assay Against a Diverse Panel of Human Tumor Cell Lines. *JNCI Journal of the National Cancer Institute*, 82(13), pp.1113-1117.
- 16. Mosaddegh, M., Ostad, S., Naghibi, F. and Hamzeloo Moghadam, M. (2006). Cytoxic effects of five species of Inula against some tumor celllines. *Iranian Journal of Pharmaceutical Research*, 2(4), pp.203-208.
- Mosaddegh, M., HamzelooMoghadam, M., Ghafari, S., NaghibI, F., Ostad, S. and Read, R. (2010). Sesquiterpene lactones from Inula oculus-christi. *Nat Prod Commun.*, 5(4), pp.511-514.
- 18. Mosmann, T. (1983). Rapid colorimetric assay for cellular growth and survival: application to proliferation and cytotoxicity assays. *J Immunol Methods*, 65(1-2): 55-63.
- 19. Brown, J. (1980). A review of the genetic effects of naturally occurring flavonoids, anthraquinones and related

compounds. *Mutation Research/Reviews in Genetic Toxicology*, 75(3), pp.243-277.

20. Hirano, T., Oka, K. and Akiba, M. (1989). Antiproliferative effects of synthetic and naturally occurring flavonoids on tumor cells of the human breast carcinoma cell line. *Res. Commun. Chem. Pathol.Pharmacol.*, 64(1), pp.69-78.