

Research article

Effects of fly ash and *P. hysterophorus* on the antioxidant status of earthworm (*Eisenia fetida*)

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Key words: Fly ash, *Parthenium hysterophorus*, *Eisenia fetida* and Antioxidant enzymes.

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Abstract

Fly ash is a serious source of air pollution since it remains air borne for a long period of time and causes health hazards. *Parthenium hysterophorus* is one among the most troublesome weeds at the global level. The aim of the present study was to evaluate the effect of fly ash and parthenium weed on antioxidant status of earthworm, *Eisenia fetida*. Earthworms were allowed to grow in the mixture of cow dung: fly ash (60:40) and cow dung: parthenium (75:25) for 60 days. The biochemical markers viz. catalase (CAT), superoxide dismutase (SOD), glutathione peroxidase (GPx) and malondialdehyde (MDA) level were measured at day 15, 30, 45 and 60. The results revealed increased MDA level, while SOD, GPx and CAT activities showed variation in both the treatments. The study indicated that fly ash and *Parthenium hysterophorus* has adverse biological effects on the model organism *Eisenia fetida*.

Introduction

Large scale industrialization, urbanization and population growth have affected the healthy relationship between man and nature. Various human activities generate huge quantities of solid wastes throughout the world and their management has become a technical and ecological challenge for all. Most of the wastes are disposed in ecologically unsustainable manner by open dumping or burning [1]. Production of large quantities of organic waste all over the world poses major environmental (offensive odors, contamination of ground water and soil) and disposal problems [2].

Fly ash, a resultant of combustion of coal at high temperature, has been regarded as a problematic solid waste all over the world [3]. A massive amount of fly ash (4750 million tons) is generated worldwide from coal-based thermal power plants [4]. Recently it has started receiving alarming attention due to its hazardous nature, wide spread usage, and the manner of disposal; leading to severe environmental pollution [5, 6]. India generates higher amount of fly ash and utilizes lower percentage of fly ash compared to other countries. Therefore, major portion of it is disposed in ash ponds near the power plants occupying more than 65,000 acres of land [7, 8]. These ash ponds have become a potential source for contamination of soil and water streams [7, 9, 10]. Leaching and accumulation of organic and inorganic toxic compounds from fly ash is of major environmental concern and known to have severe adverse impact such as bioaccumulation of metals, oxidative stress, DNA

damage and reproduction on terrestrial and aquatic ecosystems [8, 11-13].

Parthenium hysterophorus is one of the top ten worst weeds of the world and has been listed in the global invasive species database [14]. *P. hysterophorus* is an invasive alien weed of global significance [15]. The sesquiterpene lactones namely parthenin and coronopilin present in the trichomes of leaves and stems of *Parthenium*, are responsible for causing various allergies like contact dermatitis, hay fever, asthma and bronchitis in human beings [15, 16]. *P. hysterophorus* is poisonous to livestock when it is consumed or repeatedly in contact with the weed. Those animals can encounter death, rashes on their body and udders, alopecia, loss of skin pigmentation, allergic skin reactions, dermatitis, diarrhoea, anorexia and pruritus [17]. Several control methods (chemical, biological, mechanical and integrated) are being used and most of the methods are not successful due to rapid re-infestation of the plant [18]. Earthworms are universally employed as an ecosystem indicator species in ecotoxicological studies on soil contaminants [19-21]. Earthworms are sensitive, readily available and easy to handle, and chronological data are existing from their use in toxicity investigations [22]. Environmental conditions greatly affect their vital processes and the diversity of living organisms [23]. Biochemical responses in organisms against environmental stress are regarded as early warning indices of pollution in the environment. Antioxidant enzymes protect the cells from various reactive oxygen species (ROS) and hence considered as biomarkers for

assessing the environmental impact of contaminants. Enzymes like superoxide dismutase (SOD), catalase (CAT), glutathione peroxidase (GPx), glutathione reductase (GR), glutathione S-transferase (GST), as well as lipid peroxidation (LPO) [24-28] have been studied as biomarkers of environmental pollution. There is lack of data on biochemical responses in *E. fetida* by *P. hysterothorus* and fly ash.

Therefore, the aim of the present study was to understand the biological effects of fly ash and *P.hysterothorus* on the earthworm, *Eisenia fetida* and to provide additional information on their toxicological effects.

Materials and methods

Collection of Cow dung

Cow dung was obtained from nearby village.

Collection of Fly ash

Fly ash was procured from a thermal power plant, Rajpura.

Collection of *P. hysterothorus*

Parthenium hysterothorus was collected from fallow lands of Patiala.

Procurement of Earthworms

The earthworms (*Eisenia fetida*) were obtained from Punjab State Council for Science and Technology, Chandigarh.

Experimental Setup

The experiments were conducted in plastic trays, each with a capacity of 1 kg, with a hole at the bottom. The cow dung, fly ash and *P.hysterothorus* were mixed in different ratios as a bedding material:

Cow dung: Fly ash - 60:40

Cow dung: *P.hysterothorus* - 75:25

Total four plastic trays were taken. Thirty healthy earthworms were introduced in plastic trays; water was sprinkled daily on the trays using sprayer to maintain the moisture level of 55-60%. The plastic trays were kept under shade and covered with the gunny bags to avoid direct sunlight.

Biochemical Analysis

Eight earthworms were removed from each group at the interval of 15, 30, 45 and 60 days of exposure, rinsed with distilled water and kept for 48h on moist filter paper in petridishes to deplete their gut content. The earthworms were homogenized in potassium phosphate buffer (0.1M) and centrifuged at 10,000 rpm for 10 min at 4°C. The enzyme assays were performed using dual beam

UV-visible spectrophotometer from Labtronics (LT-2900).

Lipid peroxidation was measured as malondialdehyde (MDA) a thiobarbutaric acid reacting substance, using the method of Wilbur [29]. The reaction mixture contained 0.8 M HCl, 12.5% TCA and 1.23% TBA. The mixture was incubated for 15 minutes at 90°C, after cooling, reaction was measured at 530nm. Values are expressed in $\mu\text{mol mg}^{-1}$ protein using extinction coefficient of $1.56 \times 10^5 \text{ cm}^{-2} \text{ mol}^{-1}$.

SOD activity was determined as described by Das [30] in a reaction mixture containing phosphate buffer (pH 7.4), 20 mM α -methionine, 100mM hydroxylamine hydrochloride, 50 μ M EDTA, 1% triton X- 100, 100 μ M riboflavin, 0.1% naphthylethylene diamine (NED), 1.0% sulphaniamide, 5% orthophosphoric acid and 100 μ l sample. The activity was measured at 543nm.

Catalase (CAT) was estimated by the method of Aebi [31]. The reaction mixture contained 0.1mM phosphate buffer (pH 7.4) and 30 M H₂O₂ and 50 μ l sample. The rate of decomposition of H₂O₂ was measured at 240nm.

GPx activity was measured as described by Rotruck [32]. The reaction mixture contained 0.4 M Tris buffer (pH 7.0), 10 mM sodium azide, 2.5 mM hydrogen peroxide, 4mM reduced glutathione, 10% TCA, 0.3M Phosphate solution, 4mM EDTA, 0.04% Ellman's reagent, 1mM reduced glutathione and 100 μ l sample. The activity of glutathione was measured at 420nm.

Statistical analysis

The data was analyzed by Student's *t*-test using graph pad and considering $p < 0.001$ as significant and $p > 0.05$ as non significant.

Results and discussion

Results

The effects of fly ash and *P. hysterothorus* on the biochemical responses of *E. fetida* during vermicomposting are shown in Figure 1, 2, 3 and 4. The MDA level showed significant increase at 15, 30, 45 and 60 day as compared to control in fly ash groups. In *P. hysterothorus* treated groups, the same trend was observed i.e. increased MDA level at all intervals in comparison to control.

The SOD activity significantly increased at 15 and 30 day, while the level of SOD was decreased at 45 and 60 day in fly ash and *P. hysterothorus* treated groups as compared to control.

The activities of CAT showed significant elevation at 15 and 30 day, while significant reduction was observed at 45 and 60 day in fly ash treated groups as compared to control. On exposure to *P. hysterothorus*, the same trend was observed i.e. significant increase at 15 and 30 day exposure, while decline in activity of CAT was observed at 45 and 60 day in comparison to control.

A significant increase in Gpx activity was observed on 15 and 30 day, while a significant decrease was noted on 45 and 60 day as compared to control in fly ash and *P. hysterothorus* treated groups.

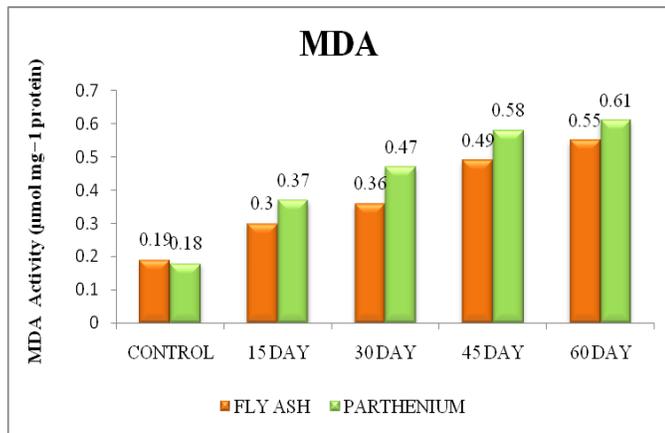


Figure 1. Effect of fly ash and *P.hysterothorus* on the MDA content of *E. fetida*.

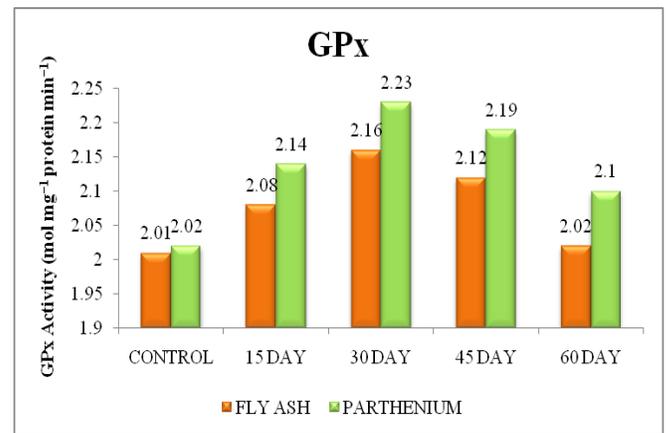


Figure 4. Effect of fly ash and *P.hysterothorus* on the Gpx activity of *E. fetida*.

Discussion

Biological molecular markers are regarded as fast, diagnostic and prognostic early warning system to detect and assess the environmental impact of wide range of contaminants, which cannot be achieved from chemical analysis of environmental samples [26, 33, 34].

Various contaminants like metals are known to induce lipid peroxidation through the formation of ROS and earthworms are particularly susceptible to peroxidation of lipids due to high content of polyunsaturated fatty acids [35, 36]. The MDA is an oxidized product of cellular lipid membranes and could be used as a sensitive biomarker of cell injury [23, 36]. In the present study, MDA levels were found to be increased in *E.fetida* after exposure to fly ash and *P.hysterothorus* at all intervals, indicating the formation of ROS. These results are in confirmation with the findings of Saint-Denis [36] and Ferreira-Cravo [28] who exposed *Eisenia fetida andrei* and polychaeta *Laeneris acuta* to Pb and Cu. Similar results were reported in *Dichogaster curgensis* exposed to fly ash [5].

The enzyme activities varied with duration of exposure. Superoxide dismutase catalyzes dismutation of superoxide anion into O₂ and H₂O₂ [26]. It is a primary remover of O₂⁻ radical and can play an important role in defending against accumulation of toxic activated oxygen species (AOS). The enhanced SOD activity resulted in increased H₂O₂ [37]. In present study, SOD activity showed elevation at day 15 and 30 as compared to control indicated that the earthworms exposed to fly ash and *P.hysterothorus* may induce the formation of O₂⁻ that resulted in the synthesis of superoxide dismutase to protect the cells from oxidant damage [38]. These results are in accordance with the findings of the study done by Łaszczycza [27] in which the earthworms were exposed to heavy metal contaminated soils. The SOD activity was decreased on day 45 and 60 in fly ash and *P.hysterothorus* treated groups. This may be due to the removal of high reactive superoxide or inactivation of

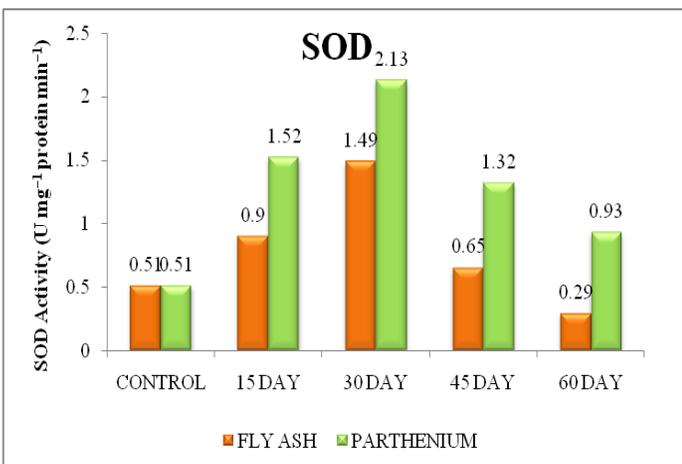


Figure 2. Effect of fly ash and *P.hysterothorus* on the SOD activity of *E. fetida*.

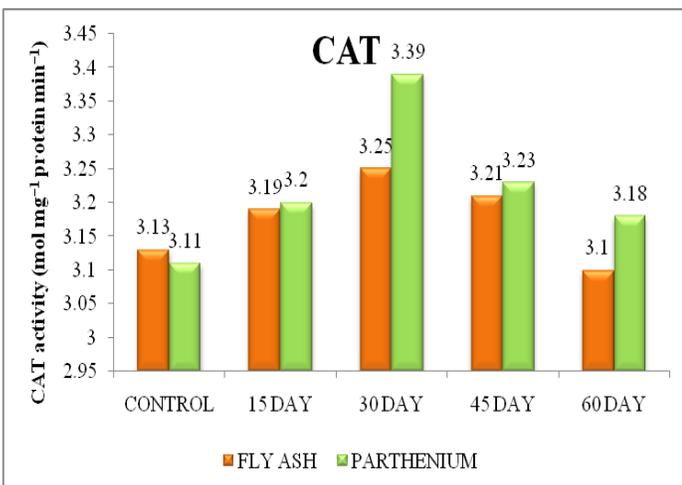


Figure 3. Effect of fly ash and *P.hysterothorus* on the CAT activity of *E. fetida*.

SOD by singlet oxygen, hydrogen peroxide and peroxy radicals [39-44].

Catalase protects the cells by eliminating H₂O₂ [26]. In the present study, the catalase activity showed significant elevation at day 15 and 30 while significant reduction was observed at 45 and 60 day when treated with fly ash and *P.hysterophorus*. The changes in catalase activity were consistent with the activity of superoxide dismutase. The catalase elevated activity at 15 and 30 day of treatment is may be due to increase in the substrate concentration produced to maintain the hydrogen peroxide level [45]. Thus, an elevation in catalase activity may be an indicator to the oxidative stress [46]. The reason for the reduction of CAT activity at 45 and 60 day exposure might be due high cellular stress or the presence of high levels of ROS [47]. Ribera [39] used CAT as a biomarker for the study of biochemical responses in earthworm (*E. fetida andrei*) exposed to carbaryl.

Glutathione peroxidase eliminates H₂O₂ by using reduced glutathione as a hydrogen donor, while glutathione reductase reduces oxidized glutathione to maintain the cellular antioxidant status. The increase in GPx activity may be a compensatory mechanism between GPx and GR [27] or NADPH availability and/or inactivation of GR by binding of metals to biomolecules [37]. The GPx activity was found to be increased at 15 and 30 day in fly ash and *P.hysterophorus* treated groups might be due to the increased demand for organism to manage peroxidative damage by reducing the level of peroxides [48]. The reduced GPx activity at 45 and 60 day may be due to some detoxification and phenotypic adaptive mechanisms [49].

Conclusion

The study reports on the effects of fly ash and *P. hysterophorus* on the antioxidant status of earthworm (*Eisenia fetida*). Toxicological effects of fly ash and parthenium weed on *E. fetida* were assessed by using biomarkers. During management and risk assessment of fly ash and parthenium weed it was found that fly ash and parthenium induces harmful effects such as oxidative stress to earthworm *E. fetida*. Therefore, use of fly ash as soil amendment and *P. hysterophorus* as feed may lead to harmful environmental hazards.

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References

1. Elvira C, Dominguez J, Sampedro L and Mato S: Vermicomposting for the paper-pulp industry. *Biocycle* 1995; 36 (6): 62–63.
2. Edwards CA and Bateer JE: The use of earthworm in environmental management. *Soil Biology and Biochemistry* 1992; 24: 1683–1689.
3. Sarojini S, Annanthakrishnasamy G, Manimegala G, Prakash M and Gunasekaran G: Effect of lignite fly ash on the growth and reproduction of Earthworm *Eisenia fetida*. *E- Journal of Chemistry* 2009; 6(2): 511–517.
4. Yao ZT, Ji XS, Sarker PK, Tang JH, Ge LQ, Xia MS and Xi YQ: A comprehensive review on the applications of coal fly ash. *Earth Science Reviews* 2015; 141: 105–121.
5. Markad VL, Kodam KM and Ghole VS: Effect of fly ash on biochemical responses and DNA damage in earthworm *Dichogaster curgensis*. *Journal of Hazardous Material* 2012; 215–216: 191–198.
6. Maity S, Bhattacharya S and Chaudhury S: Metallothionein in response in earthworms *Lampito mauritii* (Kinberg) exposed to fly ash. *Chemosphere* 2009; 77: 319–324.
7. Pandey VC, Singh JS, Singh RP, Singh N and Yunus M: Arsenic hazards in coal fly ash and its fate in Indian scenario. *Resources Conservation Recycling* 2011; 55:819–835.
8. Pandey VC and Singh N: Impact of fly ash in incorporation in soil systems. *Agriculture Ecosystem and Environment* 2010; 136:16–27.
9. Dragović S, Čujić M, Slavković - Bešković L, Gajić B, Bajat B, Kilibarda M and Onjia A: Trace element distribution in surface soils from a coal burning power production area: a case study from the largest power plant site in Serbia. *Catena* 2013; 104: 288–296.
10. Mandal A and Sengupta D: An assessment of soil contamination due to heavy metals around a coal fired thermal power plant in India. *Environmental Geology* 2006; 51: 409–420.
11. Grumiaux F, Demuyneck S, Schikorski D, Lemièrre S and Leprêtre A: Assessing the effects of FBC ash treatments of metal contaminated soils using life history traits and metal bioaccumulation analysis of earthworm *Eisenia andrei*. *Chemosphere* 2010; 79: 156–161.
12. Chakraborty R and Mukherjee A: Mutagenicity and genotoxicity of coal fly ash water leachate. *Ecotoxicology and Environmental Safety* 2009; 72: 838–842.
13. Ali M, Parvez S, Pandey S, Atif F, Kaur M, Rehman H and Raisuddin S: Fly ash leachate induces oxidative stress in freshwater fish *Channa punctata* (Bloch). *Environment International* 2004; 30: 993–998.
14. Callaway RM and Ridenour WM: Novel weapons: invasive success and the evolution of increased competitive ability. *Frontiers in Ecology and the Environment* 2004; 2(8): 436-443.
15. Kapoor RT: Awareness related survey of an invasive alien weed, *Parthenium hysterophorus* L. in Gautam Budh Nagar district, Uttar Pradesh, India. *Journal of Agricultural Technology* 2012; 8(3): 1129-1140.
16. Wiesner M, Taye T, Hoffmann A, Wilfried P, Buettner P, Buettner C, Mewis J and Ulrichs C: Impact of the pan-tropical weed *Parthenium hysterophorus* L. on human health in Ethiopia, Utilization of diversity in land use systems: Sustainable and organic approaches to meethuman needs., Tropentag, Witzhausen 2007.
17. Bezuneh TT: Phytochemistry and antimicrobial activity of *Parthenium hysterophorus* L.: a review. *Science Journal of Analytical Chemistry* 2015; 3(3):30–38.
18. Yadav A and Garg VK: Vermiconversion of biogas plant slurry and parthenium weed mixture to manure. *International Journal of Recycling of Organic Waste in Agriculture* 2016; 5:301-309.
19. Reinecke SA and Reinecke AJ: The Comet assay as biomarker of heavy metal genotoxicity in earthworms. *Archives of Environmental Contamination and Toxicology* 2004; 46:208–215.
20. Langdon CJ, Pearce TG, Meharg AA and Semple KT: Survival and behavior of the earthworms *Lumbricus rubellus* and *Dendrodrilus rubidus* from arsenate contaminated and non-contaminated sites. *Soil Biology and Biochemistry* 2001; 33:1239–1244.
21. Marino F and Morgan AJ: The time-course of metal (Ca, Cd, Cu, Pb, Zn) accumulation from a contaminated soil by three populations of the earthworm *Lumbricus rubellus*. *Applied Soil Ecology* 1999; 12:169–177.
22. OECD. Guideline for testing of chemicals: In: Earthworm acute toxicity tests, No. 207. Organization for Economic Cooperation and Development, Paris.1984.
23. Eida MF, Matter IA and Zaher FHA: Isolation and Characterization of cellulose and hemicellulolytic fungi from salt affected soils and compost. *Journal of Innovations in Pharmaceutical and Biological Sciences* 2016; 3(4): 164-170.

24. Livingstone DR, Garcia-Martinez P, Michel X, Narbonne JF, O'Hara S, Ribera D and Winston G: Oxyradical production as a pollution-mediated mechanism of toxicity in the common mussel *Mytilus edulis* and other mollusks. *Functional Ecology* 1990; 4: 415–424.
25. Dallinger R: Strategies of metal detoxification in terrestrial invertebrates: In: Dallinger R, Rainbow PS (eds) *Ecotoxicology of metals in invertebrates*. Lewis Publishers, London. 1993; 246–281.
26. Saint-Denis M, Labrot F, Narbonne JF and Ribera D: Glutathione, glutathione related enzymes, and catalase in the earthworm *Eisenia fetida* Andrei. *Archives of Environmental Contamination and Toxicology* 1998; 35:602–614.
27. Łaszczycza P, Augustyniak M, Babczyńska A, Bednarska K, Kafel A, Migula P, Wilczek G and Witas L: Profiles of enzymatic activity in earthworms from zinc lead and cadmium polluted areas near Olkusz (Poland). *Environmental International* 2004; 30: 901–910.
28. Ferreira-Cravo M, Ventura-Lima J, Sandrini JZ, Amado LL, Geracitano LA, Rebelo M, Bianchini A and Monserrat JM: Antioxidant responses in different body regions of the polychaeta *Laeonereis acuta* (Nereididae) exposed to copper. *Ecotoxicology and Environmental Safety* 2009; 72: 388–393.
29. Wilbur KM, Bernheim F and Shapiro OW: The thiobarbituric acid reagent as a test for the oxidation of unsaturated fatty acid by various agents. *Acta Biochemistry and Biophysics* 1949; 24: 305-313.
30. Das K, Samanta L and Chainy GBN: A modified spectrophotometric assay for superoxide dismutase using nitrite formation by superoxide radicals. *Indian Journal of Biochemistry and Biophysics* 2000; 37: 201-204.
31. Aebi H.E: Catalase. In: *Methods of enzymatic analysis*. Bergmeyer H.U.(ed.).Verlag Chemie. Weinheim 1983; 3: 273-286.
32. Rotruck JT, Pope AL, Ganther HE, Swanson AB, Hafeman DG and Hoekstra WG: Selenium: biochemical role as a component of glutathione peroxidase. *Science* 1973; 179: 588-590.
33. Lourenco JI, Pereira RO, Silva AC, Morgado JM, Carvalho FP, Oliveira JM, Malta MP, Paiva AA, Mendo SA and Goncalves FJ: Genotoxic endpoints in the earthworms/sub-lethal assay to evaluate natural soils contaminated by metals and radionuclides. *Journal of Hazardous Material* 2011; 186:788–795.
34. Livingstone DR: Review biotechnology and pollution monitoring: use of molecular biomarkers in the aquatic environment. *Journal of Chemistry Technology and Biotechnology* 1993; 57:195–211.
35. Krauss M, Wilcke W and Zech W: Availability of polycyclic aromatic hydrocarbons (PAHs) and polychlorinated biphenyls (PCBs) to earthworms in urban soils. *Environmental Science and Technology* 2000; 34: 4335–4340.
36. Saint-Denis M, Narbonne JF, Arnaud C and Ribera D: Biochemical responses of the earthworm *Eisenia fetida andrei* exposed to contaminated artificial soil: effects of lead acetate. *Soil Biology and Biochemistry* 2001; 33:395–404.
37. Lin D, Zhou Q, Xie X and Liu Y: Potential biochemical and genetic toxicity of triclosan as an emerging pollutant on earthworms (*Eisenia fetida*). *Chemosphere* 2010; 18:1328-1333.
38. Zhang W, Zhang M, An S, Xiong B, Li H, Cui CZ and Lin KF: Ecotoxicological effects of decabromodiphenyl ether and cadmium contamination on soil microbes and enzymes. *Ecotoxicology and Environmental Safety* 2012; 82:71-79.
39. Hodgson E K and Fridovich I: The interaction of bovine erythrocyte superoxide dismutase with hydrogen peroxide: inactivation of the enzyme. *Biochemistry* 1975; 14: 5294–5299.
40. Escobar JA, Rubio MA and Lissi EA: SOD and catalase inactivation by singlet oxygen and peroxy radicals. *Free Radical Biology and Medicine* 1996; 20:285–290.
41. Ribera D, Narbonne JF, Arnaud C and Saint-Denis M: Biochemical responses of the earthworm *Eisenia fetida andrei* exposed to contaminated artificial soil, effects of carbaryl. *Soil Biology and Biochemistry* 2001; 33:1123–1130.
42. Company R, Serafim A, Bebianno MJ, Cosson R, Shillito B and Fiala-Medioni A: Effect of cadmium, copper and mercury on antioxidant enzyme activities and lipid peroxidation in the gills of the hydrothermal vent mussel *Bathymodiolus azoricus*. *Marine Environmental Research* 2004; 58:377–381.
43. Fatima M, Mandiki SNM, Douxfils J, Silvestre, F, Coppe P and Kestemont P: Combined effects of herbicides on biomarkers reflecting immune-endocrine interactions in goldfish immune and antioxidant effects. *Aquatic Toxicology* 2007; 81:159–167.
44. Sandrini JZ, Lima JV, Regoli F, Fattorini D, Notti A, Marins LF, Monserrat JM and Kapoor RT: Antioxidant responses in the nereidid *Laeonereis acuta* (Annelida, Polychaeta) after cadmium exposure. *Ecotoxicology and Environmental Safety* 2008; 70:115–120.
45. Liu S, Zhou Q X, and Wang Y Y: Ecotoxicological responses of the earthworm *Eisenia fetida* exposed to soil contaminated with HHCb. 2011; *Chemosphere* 83:1080–86.
46. Yang X, Yufang S, Ackland ML, Yang L and Xiufeng C: Biochemical Responses of Earthworm *Eisenia fetida* Exposed to Cadmium-Contaminated Soil with Long Duration. *Bulletine of Environmental Contamination and Toxicology* 2012; 89:1148–1153.
47. Rajiv P, Rajeshwari S and Venkatesh R: Impact of *Parthenium* weeds on earthworms (*Eudrilus eugeniae*) during vermicomposting. *Environmental Science and Pollution Research* 2014; 21:12364-12371.
48. Elumalai M, Antunes C and Guilhaermeno L: Enzymatic biomarkers in the crab *Carcinus maenas* from Minho River estuary (NW Portugal) exposed to zinc and mercury. *Chemosphere* 2007; 66(7): 1249–1255.
49. Posthuma L and Van Straalen M.M: Heavy metal adaptation in terrestrial invertebrates: a review of occurrence, genetics, physiology and ecological consequences. *Comparative Biochemistry and Physiology C* 1993; 100:11-38.